

Neonicotinoid Concentrations in Arable Soils After Seed Treatment Applications in Preceding Years

Author's Name: Ainsley Jones, Paul Harrington and Gordon Turnbull

Corresponding Author:

Ainsley Jones

Food and Environment Research Agency, Sand Hutton, York YO41 1LZ

Telephone: 01904 462451

Fax: 01904 462111

e-mail: ainsley.jones@fera.gsi.gov.uk

Abstract

BACKGROUND: Concentrations of the neonicotinoid insecticides clothianidin, thiamethoxam and imidacloprid were determined in arable soils from a variety of locations in England

RESULTS: In soil samples taken from the central area of fields, concentrations of clothianidin ranged from 0.02 µg/kg to 13.6 µg/kg. Thiamethoxam concentrations were between <0.02 µg/kg and 1.50 µg/kg, imidacloprid concentrations were between <0.09 µg/kg and 10.7 µg/kg. Concentrations of clothianidin and thiamethoxam were lower in soil samples taken from the edges of fields than from the centres of fields but this difference was less pronounced for imidacloprid.

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CONCLUSION: This work gives a clear indication of the levels of neonicotinoids in arable soils after typical use of these compounds as seed dressings in the UK. There was evidence that imidacloprid was more persistent in the soils studied than clothianidin and thiamethoxam. Since clothianidin and thiamethoxam have largely superseded imidacloprid in the UK, neonicotinoid levels were lower than suggested by predictions based on imidacloprid alone.

Key words: soil; neonicotinoids; clothianidin; thiamethoxam; imidacloprid; honeybee; *Apis mellifera*; bumblebee; pollinator

1. INTRODUCTION

There has been considerable interest in recent years in the possible effects of neonicotinoid insecticides on honeybees and other pollinators. Neonicotinoids are water-soluble systemic insecticides that can be applied as seed dressings, therefore avoiding the need for spray applications. Their systemic nature means that they are carried to all parts of the plant but this can lead to contamination of nectar and pollen¹ which can, in turn, lead to exposure of pollinators. Blacquiere *et al*² outlined a number of laboratory studies describing effects of neonicotinoids on foraging and learning abilities of bees. Concern has further increased in recent years as a result of studies describing effects of neonicotinoids on foraging abilities of honeybees (*Apis mellifera*) and bumblebees^{3,4,5} and on colony development and queen production in bumblebees.⁶ As a result of the growing body of evidence, the European Food Safety Authority (EFSA) re-assessed the risk to bees and other pollinators from the three neonicotinoids of greatest concern: clothianidin, thiamethoxam and imidacloprid.^{7,8,9} As well as the reported adverse effect on bees, one of the reasons for concern around neonicotinoids are their reported persistence in soil e.g. clothianidin has a reported half life of 148 to 1,155 days in aerobic soil.¹⁰ Therefore soil-dwelling insects may be exposed to neonicotinoids for extended periods and pollinators may be exposed via contamination of

pollen and nectar from subsequent crops. There is limited published information on levels of neonicotinoids in arable soil from fields in which seed treatments had been applied in previous years and a recent review suggested that further data were required.¹¹ Krupke *et al*¹² determined neonicotinoid levels in soil from fields in which treated maize seed had been applied the previous year. It was not clear whether the levels detected arose from the treated seed or from deposition of contaminated talc that had been added to ensure a smooth flow of treated seed during planting. Most of the rest of the available data relate only to imidacloprid, the use of which has declined markedly in recent years whilst use of clothianidin and thiamethoxam has been increasing.¹¹ Schmuck *et al*¹³ detected imidacloprid levels of 12.7-17.8 µg/kg in soil samples taken from 2 sites at 7-8 months after application of cereals treated with imidacloprid treated seed. Bonmartin *et al*¹⁴ found levels between 1 µg/kg and >100 µg/kg of imidacloprid in soils from a range of fields across France in 2005. It has recently been speculated that current levels of clothianidin, thiamethoxam and imidacloprid in soils are likely to be higher than this due to their widespread use and potential to accumulate.¹¹

Neonicotinoids are also mobile in soil¹¹ so that pollinators may be exposed via plants growing in field margins if there is significant lateral movement of the pesticide.

In order to provide information on which to assess the risk to both soil-dwelling arthropods and the risk to pollinators via uptake in subsequent crops and plants in field margins we conducted some preliminary investigation to determine levels of clothianidin, thiamethoxam and imidacloprid in soil samples, taken from the central area, and from the edge, of a number of fields in Eastern England to which seeds treated with clothianidin, thiamethoxam and imidacloprid had been applied in the previous 3 years.

2. EXPERIMENTAL METHODS

2.1. Site selection and soil sampling

This was a preliminary investigation designed to obtain data in a short timescale. The time available did not allow for the design of a systematic sampling regime taking into account such factors as soil type. Instead we contacted farmers across Eastern England with whom we had previously contact and who we believed had used neonicotinoid-treated seed dressing in recent years. Since the priority was to estimate the range of concentrations across a wide area, rather than obtain highly accurate measures at each site, and due to the need to obtain samples quickly, we decided that replication of sites was more important than replication of sampling within each site. We eventually selected 18 sites that were reasonably well spread across the geographic range of interest. Only two soil samples were to be taken from each field, to a depth of 10 – 15 cm using a 25 mm diameter corer. One of these was taken from the main body of the field, the second from an area of field margin, not normally in cultivation. Sampling areas were targeted to avoid, as far as was possible, vegetation in the sample. Where possible the “Field Edge” sample was taken down-slope to increase the likelihood that groundwater from the field had percolated through the area. Fields were sampled during Spring 2013. Each soil core was transferred to a freezer and stored at approximately -18°C prior to analysis. The farmer was asked to complete a questionnaire detailing application of neonicotinoid seed dressings. Pesticide application records were collected to supplement the questionnaire when these were readily available.

2.2. Chemical Analysis

Soil samples were thoroughly mixed, a portion (10 g) of each sample was shaken with methanol (60 ml) for 1 hour, the mixture was filtered and the volume of the filtrate adjusted to 100 ml with methanol. An aliquot (2 ml) of this solution was evaporated to dryness and re-dissolved in methanol+water (1+9 by volume) prior to analysis. This solution was analysed by liquid chromatography-mass spectrometry/mass spectrometry (LC-MS/MS) using an Agilent 1290 Infinity LC system interfaced to an Agilent 6490 Triple Quad Mass

Spectrometer. Chromatography was performed on a Waters Acquity UPLC HSS analytical column with a mobile phase consisting of 5 mM ammonium acetate and methanol programmed from 2% methanol at $t=0$ to 98% methanol after 8 minutes. The solvent flow rate was 0.5 ml/min. The MS was operated in positive electrospray mode. For each analyte, two channels were monitored, each consisting of a separate MS/MS transition. One of these channels was used for quantification with the other used as confirmation.

Quantification was by means of external calibration solutions prepared in extracts obtained from control sample i.e. matrix-matched calibration. Each batch of samples analysed included a control soil sample fortified at 0.004 mg/kg from which recovery values for each analyte could be calculated.

3. RESULTS

Details of soil samples obtained are given in Table 1. Where the application rate used was known this has been indicated, in other cases the maximum allowable application rate is listed.

Percentage recovery values from concurrently analysed fortified control samples were 81% and 90% for imidacloprid, 87% and 94% for clothianidin, 72% and 93% for thiamethoxam.

Limits of detection were 0.02 $\mu\text{g}/\text{kg}$ for clothianidin and thiamethoxam, 0.09 $\mu\text{g}/\text{kg}$ for imidacloprid. Clothianidin concentrations in soil from the centre of fields to which clothianidin treated cereal seed had been drilled in Autumn 2012 ranged from 1.8 $\mu\text{g}/\text{kg}$ to 13.7 $\mu\text{g}/\text{kg}$ (Table 2). However, levels of clothianidin of up to 10.3 $\mu\text{g}/\text{kg}$ were detected in soils from the centre of fields in which clothianidin or thiamethoxam (of which clothianidin is a metabolite) was last applied in Autumn 2011 and 4.5 $\mu\text{g}/\text{kg}$ of clothianidin was detected in soils from the centre of fields on which thiamethoxam was last applied in Autumn 2010.

Clothianidin was detected in one field, and thiamethoxam detected in 7 fields, to which these chemicals had not been applied in the previous 3 years. Imidacloprid was detected in soil

samples from 15 of the 18 fields sampled, at levels up to 10.7 $\mu\text{g}/\text{kg}$ in the centre of fields, even though this chemical had apparently only been used in two of these fields since 2010.

4. DISCUSSION

The results demonstrate that detectable levels of neonicotinoids are present in soil for a considerable time after use in UK conditions. In a number of fields, residues were detected of chemicals that had not apparently been used in the previous three years. These residues are unlikely to have arisen from analytical error as the procedure used, which employed LC-MS/MS detection, is highly specific. Furthermore the acquisition of data from two MS/MS channels for each chemical allowed for one of these channels to be used for confirmatory purposes. In all cases, residues detected were present in chromatograms from both channels, and there was very good quantitative agreement between channels, giving a very high level of confidence that these residue were actually present. Such residues do not necessarily arise from applications made more than three years previously. Some may have resulted from subsequent applications made to surrounding fields e.g. from dust drift since it is clear that these chemicals were in widespread use from 2010 to 2012. This may explain the detection of clothianidin in the only field which had not received clothianidin or thiamethoxam in the last 3 years. It is unlikely to be the primary reason why thiamethoxam was detected in 7 fields to which it had not been applied for over 3 years, since in all 7 fields, levels were higher in the centres than the edges. Dust drift would be expected to result in concentrations at field edges that were comparable or higher than those in field centres. It is extremely unlikely that the presence of imidacloprid residues in 13 fields which had not received applications of this chemical in the previous 3 years could have arisen from contamination from surrounding fields since imidacloprid use in the UK has been declining in recent years; amounts used dropped by around 80% between 2005 and 2010¹¹ and the application records from the fields included in this study suggest use has remained relatively low since 2010.

It is possible that, in some cases, applications of neonicotinoids were made but were not recorded in the information supplied on neonicotinoid seed dressing applications. This may be the most obvious explanation of the relatively high levels of imidacloprid detected at 'Cambridgeshire 5'. It may also partly explain the detections of clothianidin and thiamethoxam which were not related to applications in the previous 3 years. The relatively infrequent use of imidacloprid means that unrecorded applications of the chemical are also unlikely to explain the presence of imidacloprid residues in the majority of fields studied. It seems likely that many of the imidacloprid residues and some of the thiamethoxam residues detected arise from applications made more than 3 years ago.

The dissipation rate (DT_{50}) of imidacloprid from soil has been reported to vary widely, ranging from 28-1250 days¹¹ but these estimates include laboratory studies and field studies conducted under very different conditions to those pertaining in the UK e.g. in India. Results from a study conducted in the UK on winter barley seed treated with imidacloprid suggested high persistence, with estimated DT_{50} values of 1268 and 1233 days¹⁵ whilst Bonmartin *et al*¹⁴ concluded that there was a long persistence of imidacloprid in Soils from France. Some estimates of DT_{50} for imidacloprid can be made for the 2 fields to which this chemical had been applied in the previous 3 years by comparing with initial concentrations calculated by assuming that the chemical is distributed evenly in the top 10 cm of soil and that the soil bulk density for each site is 1.2. Since each soil will have been tilled a number of times since applications of imidacloprid were made, the single samples taken are probably reasonably representative of the whole field. A DT_{50} of around 500 days would explain the measured concentration in the centre of the field 'Lincolnshire 3'. In 'Yorkshire 1' the application rate is not known but, if it had been applied at 10 g/ha then a DT_{50} of around 210 days would account for the measured concentration in the centre of the field. Imidacloprid was also detected in 13 fields which had not received applications of the chemical in the previous 3

years, with a median concentration (excluding 'Cambridgeshire 5') of 1.1 $\mu\text{g}/\text{kg}$. A DT_{50} of around 440 days would result in this concentration if imidacloprid had been applied at 10 g/ha in Autumn 2009 whilst a DT_{50} of around 560 days would give rise to this concentration from an application made, at the same rate, in Autumn 2008. Overall the results suggest that imidacloprid is highly persistent in the soil in UK conditions with dissipation rates of the order of a few hundred days.

For clothianidin, the possibility of cross contamination from subsequent applications to surrounding fields cannot be discounted and estimates of dissipation rates have therefore not been made for applications prior to Autumn 2012. Clothianidin was applied to 6 fields in Autumn 2012 but there are some difficulties associated with estimating dissipation times for these fields. The application rate in 'Cambridgeshire 1' is not known but is likely to be 60-100 g/ha. Four of the fields had received applications of clothianidin or thiamethoxam in the 12 months or so prior to Autumn 2012 so some of the resulting clothianidin residue may have originated from these earlier applications. However, for 'Lincolnshire 1' and 'Lincolnshire 2' the previous applications were at much lower rates than the rates used in Autumn 2012 and are likely to have had little effect on the measured concentration. The DT_{50} values calculated for the centre of these 2 fields, plus 'Lincolnshire 3' and 'Cambridgeshire 1' (assuming an application rate of 60 g/ha) range from 37 to 78 days with a median of 65 days. The individual DT_{50} values calculated for each field should be treated with caution since only one sample was taken from the centre and the soil will not have been mixed by tillage since application. In addition, it is possible that the levels present were enhanced by cross contamination from applications made to spring-sown crops in surrounding fields, which would mean that the DT_{50} values calculated were overestimates of real values. Despite these uncertainties, the evidence from this study is that DT_{50} values for clothianidin in UK soils are lower than reported values from field studies which range from 277-1386 days¹¹. For

thiamethoxam, estimates of DT₅₀ values for applications made in Autumn 2011 range from 75 to 109 days with a median of 65 days. However, thiamethoxam was also detected in 7 fields to which it had not been applied in the previous 3 years suggesting that dissipation rates in these fields may have slower than suggested by the calculated DT₅₀ values calculated. That said, thiamethoxam levels were all below 2 µg/kg and it is clear that there is no appreciable build-up of levels of this chemical in UK soils.

The maximum concentrations we found in real use scenarios were significantly lower than the maximum concentrations of >100 µg/kg of imidacloprid found by Bonmartin *et al.*¹⁴ The reason for the much lower levels found in this study seems to be because imidacloprid is now used rather sparingly and has been largely superseded by the less persistent clothianidin and thiamethoxam.

It is not clear what concentrations would arise in the pollen and nectar of succeeding crops and wild plants grown in the fields from this study. Schmuck *et al.*¹³ found that imidacloprid levels of <1.5 µg/kg in pollen and nectar from sunflowers grown in soils containing up to 18 µg/kg, a level similar to the maximum total neonicotinoids concentration detected in this study. Given their similar persistence and water solubility it is likely that other neonicotinoids will behave similarly. Of the crops that are attractive to pollinators which are grown from seed dressed with neonicotinoids, only oilseed rape (OSR) is widely grown in the UK. If transference to pollen and nectar is similar in OSR to sunflowers then levels in succeeding crops are likely to be <1.5 µg/kg but further information is needed to confirm this. In every case, the total levels of neonicotinoids we found in soil from field edges were lower than the levels in the centre of fields. However examination of results for individual chemicals shows some interesting differences. For clothianidin and thiamethoxam, levels were much lower in the edges of most fields than in the centres, with a small number of exceptions which may be explained by sampling error or by cross contamination from

neighbouring fields. For imidacloprid, this difference was much less marked and levels in the edges of a number of fields were comparable or even higher than levels in the centres of these fields.

Table 1. Soil samples obtained from Eastern England during spring 2013

Identifier	Recent neonicotinoid seed treatments used		
	Crop	seed treatments	Application rate (g/ha)
Lincolnshire 1	wheat	clothianidin Autumn 2012	80
	OSR	thiamethoxam Autumn 2011	15
Lincolnshire 2	wheat	clothianidin Autumn 2012	60
	OSR	clothianidin Autumn 2011	19
Lincolnshire 3	wheat	clothianidin Autumn 2012	60
	OSR	imidacloprid Autumn 2011	10
Nottinghamshire 1	OSR	clothianidin Autumn 2011	24
Lincolnshire 4	wheat	clothianidin Autumn 2012	up to 100
	wheat	clothianidin Autumn 2011	79
	OSR	thiamethoxam Autumn 2010	27
Norfolk 1	OSR	thiamethoxam Autumn 2010	up to 34
Cambridgeshire 1	wheat	clothianidin Autumn 2012	up to 100
Cambridgeshire 2	OSR	thiamethoxam Autumn 2011	up to 34
Nottinghamshire 2	OSR	thiamethoxam Autumn 2011	up to 34
Nottinghamshire 3	OSR	thiamethoxam Autumn 2011	up to 34
Yorkshire 1	OSR	imidacloprid Autumn 2011	up to 12
Cambridgeshire 3	wheat	clothianidin Autumn 2011	94
	OSR	clothianidin Autumn 2010	up to 30
Cambridgeshire 4	OSR	clothianidin Autumn 2011	up to 30
Cambridgeshire 5	OSR	clothianidin Autumn 2011	up to 30
Norfolk 2	wheat	clothianidin Autumn 2012	up to 100
	sugar beet	thiamethoxam Spring 2012	up to 78
Nottinghamshire 4	OSR	clothianidin Autumn 2011	up to 30
Nottinghamshire 5	OSR	thiamethoxam Autumn 2010	up to 34
Yorkshire 2	OSR	clothianidin Autumn 2011	up to 30

Table 2. Neonicotinoid concentrations (wet weight basis) detected in soil samples

		Imidacloprid ($\mu\text{g}/\text{kg}$)	Clothianidin ($\mu\text{g}/\text{kg}$)	Thiamethoxam ($\mu\text{g}/\text{kg}$)	Total Neonicotinoids ($\mu\text{g}/\text{kg}$)
Lincolnshire 1	Centre	2.01	13.6	0.76	16.4
	Edge	< 0.09	0.30	< 0.02	0.3
Lincolnshire 2	Centre	1.40	1.78	0.24	3.42
	Edge	0.63	1.58	0.08	2.29
Lincolnshire 3	Centre	3.95	11.5	< 0.02	15.5
	Edge	1.03	1.63	< 0.02	2.66
Nottinghamshire 1	Centre	0.55	3.20	0.05	3.8
	Edge	< 0.09	0.11	< 0.02	0.11
Lincolnshire 4	Centre	1.88	4.71	0.31	6.9
	Edge	1.22	0.81	0.08	2.11
Norfolk 1	Centre	0.33	4.49	< 0.02	4.82
	Edge	0.39	4.14	< 0.02	4.53
Cambridgeshire 1	Centre	0.74	11.0	0.16	11.9
	Edge	0.15	2.48	0.08	2.71
Cambridgeshire 2	Centre	1.71	10.3	0.64	12.7
	Edge	0.19	1.21	0.11	1.51
Nottinghamshire 2	Centre	0.12	1.12	0.19	1.43
	Edge	< 0.09	< 0.02	< 0.02	
Nottinghamshire 3	Centre	0.89	1.54	1.21	3.64
	Edge	1.88	0.18	0.14	2.2
Yorkshire 1	Centre	1.36	0.02	< 0.02	1.38
	Edge	0.38	< 0.02	< 0.02	0.38
Cambridgeshire 3	Centre	1.66	9.74	0.10	11.5
	Edge	< 0.09	0.47	< 0.02	0.47
Cambridgeshire 4	Centre	0.39	0.34	0.11	0.84
	Edge	0.64	0.09	< 0.02	0.73
Cambridgeshire 5	Centre	10.7	6.69	0.90	18.3
	Edge	6.87	0.25	0.02	7.14
Norfolk 2	Centre	< 0.09	4.25	1.50	5.75
	Edge	< 0.09	1.57	0.33	1.90
Nottinghamshire 4	Centre	1.50	2.92	0.52	4.94
	Edge	0.38	0.13	0.03	0.54
Nottinghamshire 5	Centre	< 0.09	0.32	0.54	0.86
	Edge	< 0.09	0.04	< 0.02	0.04
Yorkshire 2	Centre	< 0.09	0.51	< 0.02	0.51
	Edge	< 0.09	0.09	< 0.02	0.09

Value in bold are those in which residues were detected which were not related to applications in previous three years

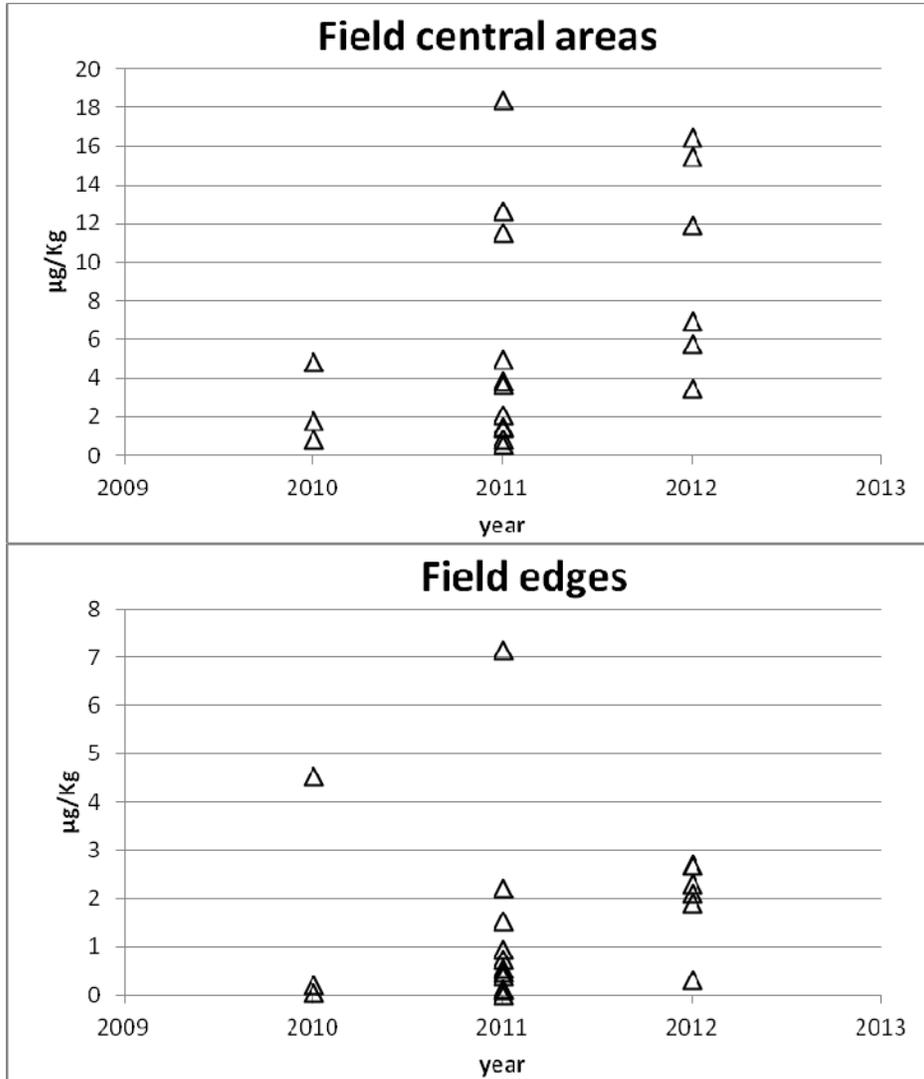


Figure 1. Total neonicotinoid concentrations ($\mu\text{g}/\text{kg}$) in soil samples from field centres and field edges plotted against last date of drilling with neonicotinoid treated seed

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